

What drives „fibrinolysis“?

R. L. Medcalf

Australian Centre for Blood Diseases, Monash University, Melbourne, Australia

Keywords

Fibrinogen, fibrin, fibrinolysis inhibitors, nervous system, plasminogen activators, plasminogen

Summary

The timely removal of blood clots and fibrin deposits is essential in the regulation of haemostasis. This is achieved by the fibrinolytic system, an enzymatic process that regulates the activation of plasminogen into its proteolytic form, plasmin. This is a self-regulated event as the very presence of fibrin initiates plasminogen activation on the fibrin surface due to the presentation of exposed C-terminal lysine residues in fibrin that allow plasminogen to position itself via its lysine binding sites and to be more efficiently cleaved by tissue-type plasminogen activator (t-PA). Hence fibrin, the ultimate substrate of plasmin during fibrinolysis, is indeed an essential cofactor in the cascade. What has now come to light is that the fibrinolytic system is not solely designed to eliminate fibrin. Indeed, it is a broad acting system that processes a variety of proteins, including many in the brain where there is no fibrin. So what drives t-PA-mediated plasminogen activation when fibrin is not available?

This review will describe the broadening role of the fibrinolytic system highlighting the importance of fibrin and other key proteins as facilitators during t-PA-mediated plasminogen activation.

Schlüsselwörter

Fibrinogen, Fibrin, Fibrinolyse-Inhibitoren, Nervensystem, Plasminogen-Aktivatoren, Plasminogen

Zusammenfassung

Die rechtzeitige Auflösung von Blutgerinnseln und Fibrinablagerungen trägt maßgeblich zur Regulierung der Hämostase bei. Erreicht wird dies durch das fibrinolytische System, einen enzymatischen Prozess, der die Aktivierung von Plasminogen zu seiner proteolytischen Form Plasmin regelt. Dies ist ein selbst reguliertes Ereignis, insofern die Plasminogen-Aktivierung auf der Fibrinoberfläche alleine durch die Anwesenheit von Fibrin, aufgrund der Präsentation von exponierten C-terminalen Lysinresten im Fibrin, ausgelöst wird; an diesen kann sich Plasminogen über seine Lysinbindungsstellen selbst anlagern und wirksamer durch den gewebespezifischen Plasminogen-Aktivator (tPA) gespalten werden. Somit ist Fibrin, das letzte Substrat des Plasmins im Verlauf der Fibrinolyse, ein wesentlicher Kofaktor in der Kaskade. Wie sich herausstellt, dient das fibrinolytische System nicht nur dazu, Fibrin zu eliminieren. Vielmehr ist es ein umfassend wirkendes System, das eine Reihe von Proteinen verarbeitet, darunter viele im Gehirn, wo es kein Fibrin gibt. Wodurch also wird die tPA-vermittelte Plasminogenaktivierung vorangetrieben, wenn kein Fibrin verfügbar ist?

In dieser Übersicht beschreiben wir die umfassendere Rolle des fibrinolytischen Systems, wobei wir vor allem die fördernde Bedeutung von Fibrin und anderen Schlüsselproteinen bei der tPA-vermittelten Plasminogen-Aktivierung beleuchten.

A short history of fibrinolysis

The dissolution of fibrin following coagulation was introduced over a century ago by a French physiologist, Albert Dastre who described the spontaneous lysis of blood clots (1). Dastre was also credited with the first introduction of the term “fibrinolysis” as reported in a 1946 review by Macfarlane and Pilling (2). The enzymatic processes underlying fibrin degradation were being actively pursued at the turn of the 20th century notably by Bertha Barker working at the Rockefeller Institute in New York (3)). Indeed, as early as 1904, a cellular basis for fibrin degradation was suggested by the Belgian scientist Hector Rulot, who reported that fibrin digestion was caused by “*leukocytes imprisoned in the meshes of fibrin*” (4). The fibrinolytic system gained a much stronger foothold in the early 1930’s through the efforts of Tillet and Garner at the Johns Hopkins Medical School (5, 6) during their description of the fibrinolytic agent produced by haemolytic streptococci that they referred to as “fibrinolysin”. Critical studies by Christensen and colleagues in New York in 1945 revealed that fibrinolysin did not degrade fibrin directly, but by activation of an unknown “zymogen or proenzyme” that existed in human plasma (7, 8). Having “plasma” as the origin of this proenzyme, Christensen named this entity as “plasminogen” and its active form as “plasmin”. However, the terms “fibrinolysin” and its pro-form “profibrinolysin” were maintained in the literature for at least another decade before they became more commonly referred to as plasmin and plasminogen, respectively. That plasma also contained an “anti-proteolytic” factor was shown as early as 1903 by Delezenne and Pozerski (9) and, as stated in the review by Macfarlane and Pilling (and being influenced by the new nomenclature introduced during this period): “This factor is probably concerned with the

Correspondence to:

Robert L. Medcalf, Ph. D.
Australian Centre for Blood Diseases
Monash University, Level 6, Burnet Building
89 Commercial Road, Melbourne 3004
Victoria, Australia
Tel. +61/3/99 03 01 33
E-mail: robert.medcalf@monash.edu

Was treibt die „Fibrinolyse“ an?

Hämostaseologie 2015; 35: 303–310
<http://dx.doi.org/10.5482/HAMO-14-10-0050>
received: October 14, 2014
accepted in revised form: December 15, 2014
epub ahead of print: January 7, 2015

absence of fibrinolysis in normal blood and may be called, for convenience, “antiplasmin” (2). The reader is also directed to a subsequent comprehensive review on the history and understanding of fibrinolysis in 1948 by MacFarlane and Biggs (10).

While the proenzyme (plasminogen) and antiplasmin had been identified relatively early, the endogenous human-derived proteases responsible for the conversion of plasminogen to plasmin were discovered later. Astrup and Permin first described what became known as tissue-type plasminogen activator (t-PA) in 1947 (11), while a “physiological activator of profibrinolysin” was identified in urine by Sobel and colleagues and named by these authors as “urokinase” (u-PA) (12).

Fibrinolysis: a current view

From the very outset, the fibrinolytic system was intrinsically linked with blood and blood clot removal and for good reason given that fibrin was the first substrate for plasmin identified. However, the forefathers of fibrinolysis could not have envisaged the extended and important role of this proteolytic system in areas far removed from fibrin removal. Nonetheless conventional fibrinolysis is still an important aspect of physiology but is beginning to take a back seat with the focus of many current research scientists turning towards these newer “non-fibrinolytic” areas.

Nowadays, the fibrinolytic system is still described in relatively straight forward terms, with the key proenzyme plasminogen still being front and centre together with t-PA and u-PA that both convert plasminogen into its potent fibrinolytic form, plasmin. However, there are many other components that will be outlined later that not only help fine-tune the fibrinolytic system, but also endow it with properties enabling this system to play important roles elsewhere in vivo.

Another important concept that will be highlighted in this review is that fibrin is not only a substrate for plasmin, it is also the initiating event of the fibrinolytic process (13, 14). This is not at all surprising, as fibrin formation under normal circumstances is only designed to provide a tem-

porary matrix, be in the guise of a blood clot or a fibrin seal during wound repair that ultimately needs to be removed in a timely manner. Mechanistically, this “self-directed” proteolysis is orchestrated by the presence of C-terminal lysine residues that are created on the fibrin surface during its formation. These lysine residues are recognised by lysine binding sites located in most of the kringle domains of plasminogen (15). This convenient arrangement not only brings plasminogen and fibrin together, but also significantly enhances the proteolytic activity of t-PA, as fibrin-bound plasminogen is activated into plasmin orders of magnitude faster in the presence of fibrin than in its absence. Hence fibrin is essentially acting as a co-factor for t-PA. To add further refinement, t-PA itself binds to fibrin (although not only via lysine residues) thereby co-locating itself juxtaposed to plasminogen on the fibrin substrate. The fibrin-selectivity of t-PA distinguishes it from u-PA, which can activate plasminogen with similar efficiency on fibrin or in solution (13).

As with all proteolytic systems, a number of check-points need to be in place not only to restrict plasmin formation, but to allow plasmin sufficient time to perform its proteolytic task. While Delezenne and Pozerski first reported the presence of an antiprotease back in 1903, plasmin formation is now known to be controlled at three distinct levels. Two of these involve specific protease inhibitors: plasminogen activator inhibitor (PAI)-1 (described in the early 1980's) (16, 17) and PAI-2 (described in 1970) (18) that target both t-PA and u-PA. As mentioned earlier, antiplasmin targets plasmin directly and is the most influential fibrinolytic inhibitor. Plasma levels of antiplasmin are very high (~1 µmol/l). This, together with its half-life of ~2.6 days (19) ensures that the half-life of plasmin activity is less than 10 ms; see (20) for review. This contrasts with the half-life of it inactive pro-form plasminogen (~2.2 days). It is also interesting to note that the association rate between antiplasmin and plasmin is the fastest described among the serine proteases and serine protease inhibitors.

The third level of plasmin regulation does not require protease inhibition at all,

but rather involves a carboxypeptidase (Thrombin activatable Fibrinolysis Inhibitor; TAFI) that functions to stabilise blood clots. TAFI enzymatically removes C-terminal lysine residues from fibrin thereby reducing the capacity of plasminogen and t-PA to bind to the fibrin surface and therefore, indirectly inhibiting fibrin breakdown (21, 22). Being activated by thrombin, TAFI became another link between coagulation and fibrinolysis, i.e. in addition to fibrin (23).

While the importance of plasminogen activation in relation to fibrinolysis is of little doubt, plasmin was also reported over 30 years ago to have important roles in the processing of other substrates, many related to tissue degradation and cancer (24). Furthermore, plasminogen activation was not restricted to the fibrin surface or to the circulation, but, importantly could be focused onto the cell surface in a manner facilitated by receptors for most of the fibrinolytic components. For example, plasminogen receptors were first described over 25 years ago and since that time, ~12 distinct cell surface plasminogen receptors have been described on various cells, nearly all containing C-terminal lysine residues (25). Similarly, receptors for u-PA (uPAR) (26) and t-PA (27, 28) have also been described. Collectively, these receptors provide a means for cell-surface localized plasmin formation where it could not only promote fibrinolysis (29), but also non-fibrinolytic functions related to cell migration (30), wound healing (31), and metastatic spread (32). It is not the purpose of this review to overview all of these numerous receptors, however their very existence underscores the fact that fibrin removal is not the only intention of the fibrinolytic system.

Thrombolysis and lysine based anti-fibrinolytics

The fibrin-dependent nature of t-PA and plasminogen provided the impetus for the pharmaceutical industry to harness this feature for the clinical development of fibrin-specific thrombolytic drugs, with t-PA being the first to reach the hospital pharmacies in the mid-1980's. This period her-

alded a mighty time for t-PA in the thrombolytic treatment of patients with myocardial infarction but this decreased in later years with the advent of angioplasty. t-PA however got an additional boost in popularity in 1995 when the FDA approved its clinical use in patients with acute ischaemic stroke (AIS) (33). Although t-PA is still today (~20 years later) the only approved drug used in AIS, there are a number of restrictions, the most critical being that it can only be administered within 4.5 hours of stroke onset (3 h in some countries, including the USA) due to risks associated with intracerebral haemorrhage and reduced net clinical benefit (34).

While the fibrin-dependency of t-PA proved beneficial for its clinical development, so too did the lysine-dependency of t-PA and plasminogen prove equally beneficial as a means of attenuating fibrinolysis. Indeed, the use of lysine analogues including epsilon-amino caproic acid (EACA) (35) and tranexamic acid (36) have been successfully exploited to limit bleeding in numerous clinical conditions and as a prophylactic treatment in some surgeries. As will be outlined in later sections, lysine-dependent plasminogen activation occurs in many other situations in the absence of fibrin raising the prospect that these treatments may have unintended consequences (see (37) for a recent review).

Non-fibrinolytic roles of the fibrinolytic system

While Albert Dastre (1) provided the first link of the fibrinolytic enzyme system with blood clots, the revolution of recombinant DNA technology and the advent of the gene knock-out mouse provided the fibrinolysis aficionados with some curious results. The t-PA and u-PA knock out (KO) mouse were also among the first KO mice to be generated. While there was an expectation that t-PA deficiency would be prothrombotic due to impaired fibrinolysis, this phenotype was surprisingly mild, with some changes in bleeding times being noted only under challenged conditions (38). The absence of a thrombotic catastrophe in mice deficient in t-PA surprised the scientific community and the field was

left pondering as to how important the fibrinolytic system actually was in controlling haemostasis with redundant systems seemingly taking up the challenge.

At about the same time, an unexpected phenotype related to t-PA deficiency was described, in an area distant to haemostasis. A series of landmark publications in the early to mid-1990's showed that t-PA deficiency had a profound phenotype in the central nervous system (CNS) (39–41). t-PA deficiency was linked with the promotion of neurotoxicity (42), learning and memory formation (43), synaptic plasticity (44), the processing of neurotrophic factors (45), the host response to addictive behaviour (46–49) and visual processing (50) just to name a few. These initial reports were published soon after the FDA approval of t-PA for AIS which sent a provocative signal to the stroke community, particularly in relation to potential consequences of t-PA-mediated neurotoxicity. The neurotoxic feature of t-PA was mechanistically linked to NMDA receptor activation (51, 52). t-PA is also involved in other CNS initiated receptor initiated events, include those associated with serotonergic (53) transmission. The finding that t-PA deficient mice were protected from direct neurotoxic challenges (42, 54), cerebral ischaemia (55) and traumatic brain injury (56, 57) clearly implicated endogenous t-PA as playing a causal role. Consistent with this brain phenotype, t-PA is widely expressed in the CNS, particularly in the hippocampus, amygdala and hypothalamus and t-PA mRNA and activity levels in the brain can be modulated pharmacologically (58). Subsequent reports implicating plasmin with t-PA activity in the CNS also begged the question as to the cellular source of plasminogen. Although initially described as an abundant liver derived plasma protein, it is now clear that many cells in the CNS can actively synthesis plasminogen (59, 60) thereby providing “brain”-t-PA with its substrate locally.

As described above, conventional fibrinolysis in the circulation has a number of well described regulatory events that restrict t-PA and plasmin activity. But how is this regulation achieved in the CNS? While PAI-1 is the most prominent t-PA inhibitor in blood, this particular inhibitor is only

weakly expressed in the CNS. Instead, the brain harbours a specific t-PA inhibitor, neuroserpin (61, 62) to control t-PA activity in this compartment. In addition, α_2 -antiplasmin is also expressed in various regions in the CNS. Curiously recent studies have suggested that α_2 -antiplasmin in the CNS may have some additional role in the CNS unrelated to plasmin inhibition (63, 64) which adds yet another dimension to the complexity of this system.

With the undeniable existence of a regulated t-PA-dependent proteolytic system in the CNS, a key question surrounds the physiological role of t-PA in this setting. Despite the fact that the early reports of the role of t-PA in the CNS tainted t-PA in a deleterious context, it certainly did not evolve in the CNS to promote harm. In recent years, this has become somewhat more topical as some reports have presented evidence that endogenous t-PA can also promote neuroprotection. t-PA was shown to inhibit zinc induced neurotoxicity back in 1999 (65), and more recently to even be protective against excitotoxic or ischaemic challenges depending on the location and concentration (66, 67). Arguably, however, a more clinically relevant role for t-PA in the CNS is its capacity to modulate blood brain barrier (BBB) permeability. This particular feature of t-PA is likely to have a bearing on its limited clinical utility in ischaemic stroke. t-PA was first shown to promote BBB permeability in a mouse model of ischaemic stroke where t-PA deficient mice displayed less parenchymal accumulation of Evans Blue dye after 6 h permanent occlusion (68). That t-PA influences permeability of the BBB (often referred to as the neurovascular unit) has been a consistent finding as reviewed in (69). This is an area of intense research activity and a number of mechanisms have been proposed to explain how t-PA opens the BBB, including interaction with LRP (a member of the low density lipoprotein receptor family), activation of matrix metalloproteinases and processing of platelet derived growth factor (PDGF)-CC and subsequent activation of the PDGFR α receptor (reviewed in (69)). Still, the fundamental question remaining is the biological purpose behind this effect of t-PA. The finding that t-PA could act to

promote glucose uptake following cerebral ischaemia (67) is consistent with a role for t-PA in regulating energy demands with BBB permeability changes complementing this effect. This however requires further exploration.

An interesting element to these effects of t-PA in the CNS is the extent to which t-PA acts alone or via plasmin (70). Plasmin certainly plays an important role in mediating some of these effects of t-PA in the CNS. Indeed, plasmin is one of the proteases (together with furin) that can convert the neurotrophic factor proBDNF into its mature form (45). t-PA-mediated neurotoxicity in vivo also requires plasmin-dependent laminin breakdown (71). While numerous other publications have indicated a requirement for plasmin generation in some of the effects of t-PA in the CNS (72), other reports have provided compelling evidence that t-PA does not require canonical (plasmin-mediated) proteolytic activation or may not require plasmin generation at all. Indeed, t-PA-mediated opening of the BBB via PDGF-CC activation mentioned above occurs in a plasmin-independent manner based on studies using plasminogen deficient mice (73). However, plasmin-dependent pathways however are not fully excluded and this could be influenced by the duration of the ischaemic event (reviewed in (70)). Other reports have indicated that t-PA can directly activate microglial cells essentially acting as a cytokine (74), and also to activate intracellular signalling pathways independently of plasmin upon binding to cell surface receptors (most likely annexin A2; (75)). The neuroprotective effect of t-PA against zinc toxicity referred to above (65) was also reported to be independent of the proteolytic actions of t-PA and was rather mediated by sequestration of zinc (76).

How can t-PA function in the absence of fibrin?

The various non-fibrinolytic effects of the plasminogen activating system are now well entrenched in the literature and have steered the field onto a number of unanticipated courses. However some fundamental questions remain. The previous 30 years of

research into the enzymatic basis of t-PA-mediated plasminogen activation in blood had one common yet important thread, that being the requirement of fibrin as an essential cofactor for t-PA to activate plasminogen. Given that the CNS is devoid of fibrin, what provides t-PA with its essential co-factor to allow it to drive plasmin formation? Alternatively, could brain t-PA operate under a different set of rules?

This still remains a pertinent question, although new research has revealed that fibrin is not the only natural cofactor for t-PA. In fact, t-PA can source its cofactor to activate plasminogen from numerous other proteins; see (77) for a review. For example, prion proteins (78, 79), beta-amyloid (80–82) and glycated proteins (83) including albumin were all shown over a decade ago to accelerate t-PA-mediated plasminogen activation, but whether there is a common determinant in these modified proteins remains to be determined. Plasminogen itself was shown to bind to dead or apoptotic cells ~100 times more effectively than live cells in a lysine-dependent manner (84), although the specific cellular binding site was not revealed.

The role of misfolded proteins as a general cofactor for t-PA (and indeed for other proteases) (85) has grown in strength in recent years. While isolated denatured/mis-

folded proteins have been shown to provide cofactor activity for t-PA, these studies did not report whether this was of any physiological relevance. Samson et al (86) reported the interesting observation that both t-PA and plasminogen both bound to the surface of dead neurons as well as other necrotic cell types, also in a lysine-dependent manner. Moreover this interaction resulted in enhanced plasmin formation and the subsequent breakdown and removal of the cell corpse. Hence, a naturally occurring non-fibrin cofactor for t-PA could indeed occur in cells to facilitate plasmin formation and the subsequent removal of the unwanted tissue. It also seems likely that plasmin-mediated turnover of necrotic tissue is associated with the well described role of plasmin(ogen) in facilitating wound repair (31, 87–89).

More detailed analysis of the role of necrotic cells in facilitating plasmin activation was reported in 2012 (90). Mass spectrophotometric analysis and enzyme kinetic studies revealed that t-PA was binding to an array of misfolded proteins formed during cell death. These proteins were mostly (but not exclusively) cytoskeletal in nature, with the most abundant of these including beta-tubulin and actin. Using a mouse model of traumatic brain injury (TBI) it was also shown that misfolded beta-tubulin

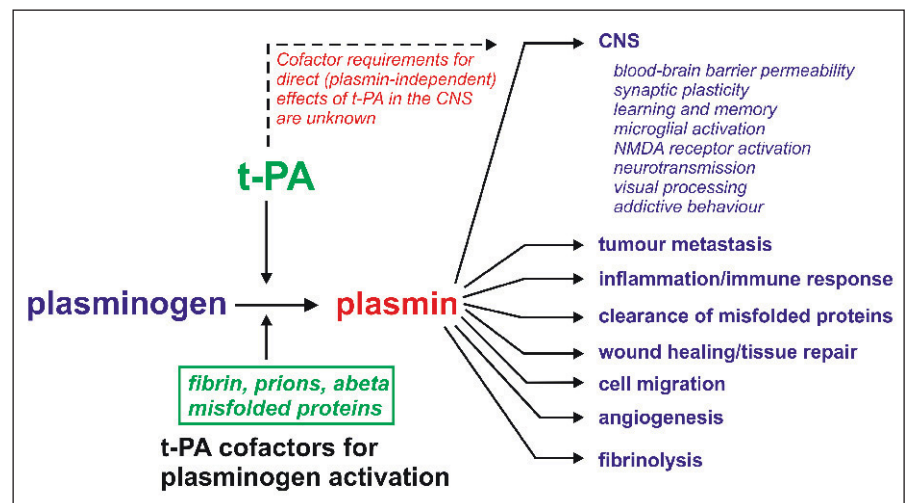


Fig. 1 Pleiotropic effects of t-PA and plasmin in physiology and in pathophysiology: Many plasmin-dependent effects of t-PA requires a specific cofactor (i.e. the classical cofactor for t-PA in fibrinolysis is fibrin) while other cofactors have also been shown to accelerate t-PA-mediated plasminogen activation in vitro (prion proteins, amyloid beta) or in vivo (misfolded/aggregated proteins). However, it remains to be determined if any of the non-fibrinolytic (plasmin-independent) effects of t-PA (mostly in relation to the CNS) requires cofactor activity.

(as determined by relative solubility) accumulated in the injured brain within 3 h of TBI and these levels subsided after ~72 h. When these experiments were performed in plasminogen deficient mice, misfolded beta-tubulin accumulated to a greater extent and persisted longer in the lesioned brain. This finding provided a strong argument for a role of the fibrinolytic system in the general removal of misfolded proteins. By extension, fibrin itself would fall under this umbrella, itself being an insoluble cross beta structured protein (77). The paper by Samson et al. (90) also described a novel means of intracellular protein aggregation occurring late following cell injury and was reliant on disulphide bonding. This novel form of aggregation was coined “nucleocytoplasmic coagulation (NCC)” (90). NCC was shown to be an injury-induced aggregation event that, akin to fibrin formation in a blood clot, encapsulates the necrotic material and subsequently facilitates their plasmin-mediated removal.

A revised definition of the fibrinolytic system?

So what is the role of the fibrinolytic system really, and what drives it? There is no dispute that this enzyme cascade can remove fibrin, although there is an element of redundancy given that fibrinolysis is largely unaltered in t-PA deficient mice. That fibrin was shown to be the first substrate for plasmin was enough to give the process its current name, but the fact that many misfolded proteins can substitute for fibrin has broadened the scope of this system to one that appears to be a general surveillance system for proteins no longer needed, or for proteins that have evolved to exist temporarily. And what of the CNS? The role of t-PA in learning and memory, synaptic plasticity, modulation of blood brain barrier permeability etc. may have different cofactor requirements or may not be needed at all, particularly in relation to any plasmin-independent effect of t-PA. A summary of the pleiotrophic effects of t-PA and the various co-factor requirements are presented (► Fig. 1). Nonetheless, the various plasmin-dependent and -independent

attributes of t-PA in the CNS has opened up a series of questions that will no doubt be addressed in the near future. Even with our current extended knowledge, it is reasonable to conclude that the original concept for the fibrinolytic system, being focused on fibrin, is a massive understatement.

Conflict of interest

The author declares that there is no conflict of interest in the content of this article

References

1. Dastre A. Arch Physiol 1893; 5: 661.
2. Macfarlane RG, Pilling J. Observations on fibrinolysis; plasminogen, plasmin, and antiplasmin content of human blood. Lancet 1946; 2: 562–565.
3. Barker BI. The enzymes of fibrin. J Exp Med 1908; 10: 343–53.
4. Rulot H. Arch Intern de Physiol 1904; i: 152.
5. Garner RL, Tillett WS. Biochemical studies on the fibrinolytic activity of hemolytic Streptococci: I. Isolation and characterization of fibrinolysin. J Exp Med 1934; 60: 239–254.
6. Tillett WS, Garner RL. The fibrinolytic activity of hemolytic Streptococci. J Exp Med 1933; 58: 485–502.
7. Christensen LR, Macleod CM. A proteolytic enzyme of serum: Characterization, activation, and reaction with inhibitors. J General Physiol 1945; 28: 559–583.
8. Christensen LR. Streptococcal fibrinolysis: A proteolytic reaction due to a serum enzyme activated by streptococcal fibrinolysin. J General Physiol 1945; 28: 363–383.
9. Delezene C, Pozerski E. 690. CR Soc Biol Paris 1903; 55: 327.
10. Macfarlane RG, Biggs R. Fibrinolysis; its mechanism and significance. Blood 1948; 3: 1167–1187.
11. Astrup T, Permin PM. Fibrinolysis in the animal organism. Nature 1947; 159: 681.
12. Sobel GW, Mohler SR, Jones, NW et al. Urokinase: an activator of plasma profibrinolysin extracted from urine. Am J Physiol 1952; 171: 768–769.
13. Cesarman-Maus G, Hajjar KA. Molecular mechanisms of fibrinolysis. Br J Haematol 2005; 129: 307–321.
14. Medcalf RL. Fibrinolysis, inflammation, and regulation of the plasminogen activating system. J Thromb Haemost 2007; 5 (Suppl 1): 132–142.
15. Law RH, Caradoc-Davies T, Cowieson N et al. The X-ray crystal structure of full-length human plasminogen. Cell reports 2012; 1: 185–190.
16. Loskutoff DJ, van Mourik JA, Erickson LA, Lawrence D. Detection of an unusually stable fibrinolytic inhibitor produced by bovine endothelial cells. Proc Natl Acad Sci USA 1983; 80: 2956–2960.
17. Coleman PL, Barouski PA, Gelehrter TD. The dexamethasone-induced inhibitor of fibrinolytic activity in hepatoma cells. A cellular product which specifically inhibits plasminogen activation. J Biol Chem 1982; 257: 4260–4264.
18. Kawano T, Morimoto K, Uemura Y. Partial purification and properties of urokinase inhibitor from human placenta. J Biochem 1970; 67: 333–342.
19. Collen D, Wiman B. Turnover of antiplasmin, the fast-acting plasmin inhibitor of plasma. Blood 1979; 53: 313–324.
20. Ellis V. The Plasminogen Activation System in Normal Tissue Remodeling. In: Behrendt N (ed). Matrix Proteases in Health and Disease: O-BK Wiley, 2012: 25–55.
21. Bajzar L, Manuel R, Nesheim ME. Purification and characterization of TAFI, a thrombin-activable fibrinolysis inhibitor. J Biol Chem 1995; 270: 14477–14484.
22. Nesheim M, Bajzar L. The discovery of TAFI. J Thromb Haemost 2005; 3: 2139–2146.
23. Bajzar L, Morser J, Nesheim M. TAFI, or plasma procarboxypeptidase B, couples the coagulation and fibrinolytic cascades through the thrombin-thrombomodulin complex. J Biol Chem 1996; 271: 16603–16608.
24. Dano K, Andreasen PA, Grondahl-Hansen J et al. Plasminogen activators, tissue degradation, and cancer. Advances in cancer research 1985; 44: 139–266.
25. Miles LA, Parmer RJ. Plasminogen receptors: the first quarter century. Semin Thromb Hemost 2013; 39: 329–337.
26. Vassalli JD, Baccino D, Belin D. A cellular binding site for the Mr 55,000 form of the human plasminogen activator, urokinase. J Cell Biol 1985; 100: 86–92.
27. Orth K, Madison EL, Gething MJ et al. Complexes of tissue-type plasminogen activator and its serpin inhibitor plasminogen-activator inhibitor type 1 are internalized by means of the low density lipoprotein receptor-related protein/alpha 2-macroglobulin receptor. Proc Natl Acad Sci USA 1992; 89: 7422–7426.
28. Ellis V, Whawell SA. Vascular smooth muscle cells potentiate plasmin generation by both urokinase and tissue plasminogen activator-dependent mechanisms: evidence for a specific tissue-type plasminogen activator receptor on these cells. Blood 1997; 90: 2312–2322.
29. Dassah M, Deora AB, He K, Hajjar KA. The endothelial cell annexin A2 system and vascular fibrinolysis. Gen Physiol Biophys 2009; 28: F20–F28.
30. Ploplis VA, French EL, Carmeliet P et al. Plasminogen deficiency differentially affects recruitment of inflammatory cell populations in mice. Blood 1998; 91: 2005–2009.
31. Shen Y, Guo Y, Mikus P et al. Plasminogen is a key proinflammatory regulator that accelerates the healing of acute and diabetic wounds. Blood 2012; 119: 5879–5887.
32. Ploplis VA, Castellino FJ. Nonfibrinolytic functions of plasminogen. Methods 2000; 21: 103–110.
33. NINDS. Tissue plasminogen activator for acute ischemic stroke. The National Institute of Neurological Disorders and Stroke rt-PA Stroke Study Group. N Engl J Med 1995; 333: 1581–1587.
34. Lees KR, Bluhmki E, von Kummer R et al. Time to treatment with intravenous alteplase and outcome in stroke: an updated pooled analysis of ECASS, ATLANTIS, NINDS, and EPITHET trials. Lancet 2010; 375: 1695–1703.

35. Yurka HG, Wissler RN, Zanghi CN et al. The effective concentration of epsilon-aminocaproic Acid for inhibition of fibrinolysis in neonatal plasma in vitro. *Anesth Analg* 2010; 111: 180–184.
36. McCormack PL. Tranexamic acid: a review of its use in the treatment of hyperfibrinolysis. *Drugs* 2012; 72: 585–617.
37. Draxler DFM, R.L. The fibrinolytic system – More than fibrinolysis? *Transfusion Med Rev* 2014; in press.
38. Carmeliet P, Schoonjans L, Kieckens L et al. Physiological consequences of loss of plasminogen activator gene function in mice. *Nature* 1994; 368: 419–424.
39. Samson AL, Medcalf RL. Tissue-type plasminogen activator: a multifaceted modulator of neurotransmission and synaptic plasticity. *Neuron* 2006; 50: 673–678.
40. Yepes M, Lawrence DA. New functions for an old enzyme: nonhemostatic roles for tissue-type plasminogen activator in the central nervous system. *Exp Biol Med* 2004; 229: 1097–1104.
41. Melchor JP, Strickland S. Tissue plasminogen activator in central nervous system physiology and pathology. *Thromb Haemost* 2005; 93: 655–660.
42. Tsirka SE, Gualandris A, Amaral DG, Strickland S. Excitotoxin-induced neuronal degeneration and seizure are mediated by tissue plasminogen activator. *Nature* 1995; 377: 340–344.
43. Baranes D, Lederfein D, Huang YY et al. Tissue plasminogen activator contributes to the late phase of LTP and to synaptic growth in the hippocampal mossy fiber pathway. *Neuron* 1998; 21: 813–825.
44. Pawlak R, Rao BS, Melchor JP et al. Tissue plasminogen activator and plasminogen mediate stress-induced decline of neuronal and cognitive functions in the mouse hippocampus. *Proc Natl Acad Sci USA* 2005; 102: 18201–18206.
45. Pang PT, Teng HK, Zaitsev E et al. Cleavage of proBDNF by tPA/plasmin is essential for long-term hippocampal plasticity. *Science* 2004; 306: 487–491.
46. Pawlak R, Melchor JP, Matys T et al. Ethanol-withdrawal seizures are controlled by tissue plasminogen activator via modulation of NR2B-containing NMDA receptors. *Proc Natl Acad Sci USA* 2005; 102: 443–448.
47. Bahi A, Kusnecov A, Dreyer JL. The role of tissue-type plasminogen activator system in amphetamine-induced conditional place preference extinction and reinstatement. *Neuropsychopharmacology* 2008; 33: 2726–2734.
48. Maiya R, Zhou Y, Norris EH et al. Tissue plasminogen activator modulates the cellular and behavioral response to cocaine. *Proc Natl Acad Sci USA* 2009; 106: 1983–1988.
49. Nagai T, Yamada K, Yoshimura M et al. The tissue plasminogen activator-plasmin system participates in the rewarding effect of morphine by regulating dopamine release. *Proc Natl Acad Sci USA* 2004; 101: 3650–3655.
50. Muller CM, Griesinger CB. Tissue plasminogen activator mediates reverse occlusion plasticity in visual cortex. *Nat Neurosci* 1998; 1: 47–53.
51. Nicole O, Docagne F, Ali C et al. The proteolytic activity of tissue-plasminogen activator enhances NMDA receptor-mediated signaling. *Nature Medicine* 2001; 7: 59–64.
52. Samson AL, Nevin ST, Croucher D et al. Tissue-type plasminogen activator requires a co-receptor to enhance NMDA receptor function. *J Neurochem* 2008; 107: 1091–1101.
53. Pothakos K, Robinson JK, Gravanis I et al. Decreased serotonin levels associated with behavioral disinhibition in tissue plasminogen activator deficient (tPA^{-/-}) mice. *Brain Res* 2010; 1326: 135–142.
54. Liberatore GT, Samson A, Bladin C et al. Vampire bat salivary plasminogen activator (desmoteplase): a unique fibrinolytic enzyme that does not promote neurodegeneration. *Stroke* 2003; 34: 537–543.
55. Nagai N, Zhao BQ, Suzuki Y et al. Tissue-type plasminogen activator has paradoxical roles in focal cerebral ischemic injury by thrombotic middle cerebral artery occlusion with mild or severe photochemical damage in mice. *J Cereb Blood Flow Metab* 2002; 22: 648–651.
56. Mori T, Wang X, Kline AE et al. Reduced cortical injury and edema in tissue plasminogen activator knockout mice after brain trauma. *NeuroReport* 2001; 12: 4117–4120.
57. Sashindranath M, Sales E, Daglas M et al. The tissue-type plasminogen activator-plasminogen activator inhibitor 1 complex promotes neurovascular injury in brain trauma: evidence from mice and humans. *Brain* 2012; 135: 3251–3264.
58. Sashindranath M, Samson AL, Downes CE et al. Compartment- and context-specific changes in tissue-type plasminogen activator (tPA) activity following brain injury and pharmacological stimulation. *Lab Invest* 2011; 91: 1079–1091.
59. Basham ME, Seeds NW. Plasminogen expression in the neonatal and adult mouse brain. *J Neurochem* 2001; 77: 318–325.
60. Taniguchi Y, Inoue N, Morita S et al. Localization of plasminogen in mouse hippocampus, cerebral cortex, and hypothalamus. *Cell and tissue research* 2011; 343: 303–317.
61. Berger P, Kozlov SV, Krueger SR, Sonderegger P. Structure of the mouse gene for the serine protease inhibitor neuroserpin (PI12). *Gene* 1998; 214: 25–33.
62. Hastings GA, Coleman TA, Haudenschild CC et al. Neuroserpin, a brain-associated inhibitor of tissue plasminogen activator is localized primarily in neurons. Implications for the regulation of motor learning and neuronal survival. *J Biol Chem* 1997; 272: 33062–33067.
63. Kawashita E, Kanno Y, Asayama H et al. Involvement of alpha2-antiplasmin in dendritic growth of hippocampal neurons. *J Neurochem* 2013; 126: 58–69.
64. Kawashita E, Kanno Y, Ikeda K et al. Altered behavior in mice with deletion of the alpha2-antiplasmin gene. *PLoS one* 2014; 9: e97947.
65. Kim YH, Park JH, Hong SH, Koh JY. Nonproteolytic neuroprotection by human recombinant tissue plasminogen activator. *Science* 1999; 284: 647–650.
66. Echeverry R, Wu J, Haile WB et al. Tissue-type plasminogen activator is a neuroprotectant in the mouse hippocampus. *J Clin Invest* 2010; 120: 2194–2205.
67. Wu F, Wu J, Nicholson AD et al. Tissue-type plasminogen activator regulates the neuronal uptake of glucose in the ischemic brain. *J Neurosci* 2012; 32: 9848–9858.
68. Yepes M, Sandkvist M, Moore EG et al. Tissue-type plasminogen activator induces opening of the blood-brain barrier via the LDL receptor-related protein. *J Clin Invest* 2003; 112: 1533–1540.
69. Schielke GL, D.A. Plasminogen Activators in Ischemic Stroke. *Matrix Proteases in Health and Disease: Wiley-VCH* 2012.
70. Niego B, Medcalf RL. Plasmin-dependent modulation of the blood-brain barrier: a major consideration during tPA-induced thrombolysis? *J Cereb Blood Flow Metab* 2014; 34: 1283–1296.
71. Chen ZL, Strickland S. Neuronal death in the hippocampus is promoted by plasmin-catalyzed degradation of laminin. *Cell* 1997; 91: 917–925.
72. Baron A, Hommet Y, Casse F, Vivien D. Tissue-type plasminogen activator induces plasmin-dependent proteolysis of intracellular neuronal nitric oxide synthase. *Biol Cell* 2010; 102: 539–547.
73. Su EJ, Fredriksson L, Geyer M et al. Activation of PDGF-CC by tissue plasminogen activator impairs blood-brain barrier integrity during ischemic stroke. *Nat Med* 2008; 14: 731–737.
74. Rogove AD, Siao C, Keyt B et al. Activation of microglia reveals a non-proteolytic cytokine function for tissue plasminogen activator in the central nervous system. *J Cell Sci* 1999; 112: 4007–4016.
75. Lee HY, Hwang IY, Im H et al. Non-proteolytic neurotrophic effects of tissue plasminogen activator on cultured mouse cerebrocortical neurons. *J Neurochem* 2007; 101: 1236–1247.
76. Siddiqi MM, Tsirka SE. Modulation of zinc toxicity by tissue plasminogen activator. *Mol Cell Neurosci* 2004; 25: 162–171.
77. Gebbink MF, Bouma B, Maas C, Bouma BN. Physiological responses to protein aggregates: fibrinolysis, coagulation and inflammation (new roles for old factors). *FEBS Lett* 2009; 583: 2691–2699.
78. Ellis V, Daniels M, Misra R, Brown DR. Plasminogen activation is stimulated by prion protein and regulated in a copper-dependent manner. *Biochemistry* 2002; 41: 6891–6896.
79. Epple G, Schleuning WD, Kettelgerdes G et al. Prion protein stimulates tissue-type plasminogen activator-mediated plasmin generation via a lysine-binding site on kringle 2. *J Thromb Haemost* 2004; 2: 962–968.
80. Kranenburg O, Gent YY, Romijn EP et al. Amyloid-beta-stimulated plasminogen activation by tissue-type plasminogen activator results in processing of neuroendocrine factors. *Neuroscience* 2005; 131: 877–886.
81. Kruijthof EK, Schleuning WD. A comparative study of amyloid-beta (1–42) as a cofactor for plasminogen activation by vampire bat plasminogen activator and recombinant human tissue-type plasminogen activator. *Thromb Haemost* 2004; 92: 559–567.
82. Kingston IB, Castro MJ, Anderson S. In vitro stimulation of tissue-type plasminogen activator by Alzheimer amyloid beta-peptide analogues. *Nat Med* 1995; 1: 138–142.
83. Bobbink IW, Tekelenburg WL, Sixma JJ et al. Glycated proteins modulate tissue-plasminogen activator-catalyzed plasminogen activation. *Biochem Biophys Res Commun* 1997; 240: 595–601.
84. O'Mullane MJ, Baker MS. Loss of cell viability dramatically elevates cell surface plasminogen binding and activation. *Exp Cell Res* 1998; 242: 153–164.

85. Maas C, Govers-Riemslog JW, Bouma B et al. Misfolded proteins activate Factor XII in humans, leading to kallikrein formation without initiating coagulation. *J Clin Invest* 2008; 118: 3208–3218.
86. Samson AL, Borg RJ, Niego B et al. A nonfibrin macromolecular cofactor for tPA-mediated plasmin generation following cellular injury. *Blood* 2009; 114: 1937–1946.
87. Wang AY, Shen Y, Wang JT et al. Animal models of chronic tympanic membrane perforation: in response to plasminogen initiates and potentiates the healing of acute and chronic tympanic membrane perforations in mice. *Clin Transl Med* 2014; 3: 5.
88. Shen Y, Guo Y, Wilczynska M et al. Plasminogen initiates and potentiates the healing of acute and chronic tympanic membrane perforations in mice. *J Transl Med* 2014; 12: 5.
89. Romer J, Bugge TH, Pyke C et al. Impaired wound healing in mice with a disrupted plasminogen gene. *Nat Med* 1996; 2: 287–292.
90. Samson AL, Knaupp AS, Sashindranath M et al. Nucleocytoplasmic coagulation: An injury-induced aggregation event that disulfide crosslinks proteins and facilitates their removal by plasmin. *Cell Rep* 2012; 2: 889–901.

Anzeige